

METHOD FOR PRODUCING CELLULOSE DERIVATIVES

INDUSTRIAL FIELD

The present invention relates to a method for producing cellulose derivatives. More specifically, it relates to a method
5 for producing cellulose derivatives from enzymatically-treated cellulose.

BACKGROUND ART

Cellulose derivatives include cellulose ethers. Cellulose ethers, such as methyl cellulose (MC), ethyl cellulose (EC),
10 hydroxyethyl cellulose (HEC), hydroxypropyl cellulose (HPC) and carboxymethyl cellulose (CMC), are water-soluble or water-suspensible white polymers which are non-caloric, odorless and tasteless. Therefore, such cellulose ethers are widely used in various fields, for example, foods, chemicals, cosmetics, paper-
15 making and fibers, as thickeners, binders, stabilizers, suspending agents etc.

With the recent development of novel uses, there is a demand cellulose ethers with high-grade properties that are applicable to such novel uses. For example, there is a demand for
20 cellulose ethers giving a reduced amount of microgel. Microgel as referred to herein is cellulose ether in the form of a semi-dissolved gel which is not completely solubilized in solvents because the etherification is incomplete.

To produce cellulose ethers, a method has heretofore been
25 employed in which a lignocellulose, such as wood pulp or linter pulp, is soaked in a strong alkali solution to give an alkali cellulose, and thereafter the resulting alkali cellulose is treated with a suitable etherifying agent (e.g., methyl chloride, ethyl chloride, ethylene oxide, propylene oxide, monochloroacetic
30 acid).

The cellulose ethers as produced according to this method could not always have satisfactory characteristics for some uses, as the properties are much influenced by the degree of substitution (degree of etherification) and the distribution of substituents. For example, cellulose ethers with a low degree of substitution are poorly soluble in solvents, because of uneven etherification. In addition, these often give a semi-dissolved gel substance which is referred to as a microgel. The gel substance, microgel is not only sensually (visually and tactually) unfavorable but also results in poor filterability of solutions of the cellulose ethers.

Enzymatic treatment of pulp has been studied, using cellulases and xylanases as the enzymes. For example, in 1986, Viikari et al. reported in Proceedings of the Symposium on Biotechnology in the Pulp and Paper Industry, 3rd International Conference, a method of enzymatically pre-treating pulp prior to bleaching it to thereby reduce the amounts of the chemicals to be used in the subsequent bleaching step.

EP 382,576 discloses a method of treating CMC with a cellulase to produce CMC hydrolysates. However, this method involves an enzymatic treatment of CMC after the etherification of pulp, so it does not reduce the amount of microgel during the etherification step. Accordingly, if the microgel is to be removed, a large amount of the enzyme must be used, resulting in noticeable reduction in the yield of the intended hydrolysates. On the other hand, if the yield of the hydrolysates is to be increased in this method, the amount of the enzyme to be used therein must be reduced. However, this is problematic in that the enzymatic treatment of the microgel is insufficient, resulting in still leaving a large amount of microgel in the system. In addition, since the viscosity of the hydrolysates obtained in this method is much lowered, as compared with that of the

original CMC, the hydrolysates are disadvantageous when used as thickeners or binders.

DE 44 40 245 C1 discloses a method of producing hydroxyalkyl-cellulose ethers by pre-treating cellulose with a cellulase solution, followed by a treatment with epoxyalkane in the presence of a quaternary ammonium base. The cellulase treatment reduced the degree of polymerization. No effect on the filtration rate is reported.

Given the situation, the object of the present invention is to provide cellulose derivatives with improved filterability while preventing the formation of microgel in the step of producing cellulose derivatives from pulp.

STATEMENT OF THE INVENTION

We, the present inventors have assiduously studied in order to solve the above-mentioned problems in the prior art, and have found that treating a pulp with a hemicellulase, e.g. a xylanase such as that derived from *Bacillus* sp. SD902, prior to chemical modification results in excellent cellulose derivatives that could not be obtained by any conventional methods. Specifically, the cellulose derivatives produced by this method have improved filterability and increased water-solubility. In addition, according to this method, the formation of microgel is minimized, and the intramolecular distribution of substituents in the cellulose derivative is made more uniform. On the basis of these findings, we have completed the present invention.

Accordingly, the invention provides a method for producing a cellulose derivative, which comprises

- a) treating the pulp with a hemicellulase, and
- b) chemically modifying the treated pulp.

Now, the present invention is described in detail hereinafter.

DETAILED DESCRIPTION OF THE INVENTION

Pulp

The pulp to be used in the present invention may be steamed or bleached pulp derived from coniferous trees (softwood), broad-
5 leaved trees (hardwood) or non-wood plants. The non-wood pulp may be pulp produced from liber plants, such as *kozo* (paper mulberry; *Broussonetia kazinoki*), *mitsumata* (*Edgeworthia papyrifera*), Manila hemp, kenaf; or from hard fiber plants, such as straw, sugar cane, bagasse. In addition, cellulose derivatives having a
10 low degree of chemical modification such as etherification, and also regenerated celluloses can also be used as the starting pulp in the present invention.

Hemicellulase

The hemicellulase used in this invention is preferably an
15 enzyme that hydrolyzes β -1,4-glycoside bonds. One enzyme or a plurality of enzymes may be used. Some preferred types of hemicellulase are xylanase, mannanase and xylo-glucanase.

The xylanase may be a xylanolytic enzyme obtained from any known source of xylanolytic enzymes. Preferably the xylanolytic
20 enzyme may be obtained from microbial sources, in particular from a filamentous fungus or yeast, or from a bacteria.

Preferred xylanolytic enzymes of fungal origin are xylanases derived from a strain of *Aspergillus*, in particular *A. aculeatus*, *A. awamori*, *A. nidulans*, *A. niger*, *A. kawachii*, or *A. tubigensis*, *Aureobasidium*, *Chaetomium*, in particular *C. gracile*, *Cochliobolus*, in particular *C. carbonum*, *Disporotrichum*, in particular *D. dimorphosporum*, *Humicola*, in particular *H. insolens*, *Neocallimastix*, in particular *N. patriciarum*, *Orpinomyces*, *Penicillium*, in particular *P. janthinellum*,
30 *Thermomyces*, in particular *T. lanuginosus* (syn. *Humicola*

lanuginosa), or *Trichoderma*, in particular *T. longibrachiatum*, or *T. reesei*.

Preferred xylanolytic enzymes of bacterial origin are xylanases derived from a strain of *Bacillus*, in particular *B. pumilus*, *B. stearothermophilus*, or *B. subtilis*, *Cellulomonas fimi*, in particular *C. fimi*, *Clostridium*, in particular *C. thermocellum*, *Dictyoglomus*, in particular *D. thermophilum*, *Microtetraspora*, in particular *M. flexuosa*, *Streptomyces*, in particular *S. olivochromogenes*, or *Thermomonospora*.

10 A particularly preferred xylanase is derived from *Bacillus* sp. SD902; it may be produced by cultivation of the strain and recovery of the xylanase as described in EP 720,649. The enzyme is hereinafter referred to as SDX enzyme. *Bacillus* sp. strain SD902 was deposited for the purposes of patenting at the National
15 Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, Ministry of International Trade and Industry, 1-3 Higashi 1-chome, Tsukuba-shi, Ibaraki-ken 305, Japan. It was deposited by Showa Denko K.K. on 25 December 1992 under deposit No. FERM P-13356, was transferred on 22
20 December 1993 to international deposit FERM BP-4508 under the terms of the Budapest Treaty, and was later assigned to Novo Nordisk A/S.

The mannanase may be obtained from microbial sources, in particular from a filamentous fungus or yeast, or from a bacteria.
25 A preferred mannanase of fungal origin is derived from *Trichoderma*, particularly *T. reesei*. This enzyme may be produced as described in WO 93/24622.

The enzyme to be used herein is not always required to be pure, but any of cell-free supernatants as obtained through
30 centrifugation of cultures of enzyme-producing cells, or crude enzyme extracts as extracted from incubated cells may be used.

The activity of the enzyme for use in the present invention can be determined by quantifying the reducing sugar as formed

through the enzymatic reaction with its substrate (e.g., xylan in the case of xylanase), at pH 7 and at 50°C, according to a method of using 3,5-dinitrosalicylic acid. One unit (U) for the enzymatic activity indicates the amount of the enzyme that forms
5 1 μ mol of reducing sugar (e.g., xylose in the case of xylanase) per minute.

Conditions for enzyme treatment

The conditions for the enzymatic treatment of pulp according to the invention are not particularly limited. The pH,
10 temperature and process time for the treatment may be suitably defined in such a manner that the enzyme being used is kept active within the defined ranges. Typical conditions are: a temperature between 20°C and 90°C, preferably between 40°C and 80°C; process time between 15 minutes and 24 hours, preferably
15 between 30 minutes and 5 hours; and a pH between 3 and 9, preferably between 4 and 8.

The amount of the enzyme to be added to pulp may be from 1 to 1000 U/g (based on pulp dry matter), preferably from 2 to 250 U/g. If the amount of the enzyme added is smaller than 1 U/g, it
20 may be too small to attain the intended enzymatic treatment; an enzyme amount larger than 1000 U/g is not preferred because the pulp yield may be reduced.

The concentration of pulp to be in the system may be any one that ensures satisfactory stirring and mixing of pulp
25 therein, but is preferably in the range 1 - 20 % by weight.

Chemical modification

The chemical modification that follows the enzymatic treatment in the method of the present invention is preferably etherification, more preferably alkyl etherification,
30 hydroxyalkyl etherification or carboxyalkyl etherification, even more preferably methyl etherification, ethyl etherification,

hydroxyethyl etherification, hydroxypropyl etherification or carboxymethyl etherification.

The method used in the chemical modification of the enzymatically-treated pulp can be any known method. For example, 5 CMC, MC, EC, HEC or HPC may be produced from pulp by two typical methods. One is an aqueous method where an aqueous medium is used as the reaction solvent; and the other is a solvent method where an organic solvent is used.

Examples of aqueous methods for producing CMC are an alkali 10 cellulose method where pulp is soaked in a solution of sodium hydroxide, and powdery sodium monochloroacetate is added thereto while beating and stirring it; and a monochlorine method where pulp is soaked in an aqueous solution of sodium monochloroacetate, and sodium hydroxide is added thereto while 15 beating and stirring it.

Examples of the solvent method are a 6-fold method that uses a mixed solvent of ethanol and benzene; and a 30-fold method that uses an aqueous solution of 2-propanol.

Apart from such chemical modification, enzymatic 20 modification may also be used.

EXAMPLES

Now, the present invention is describe in more detail with reference to the following examples that are based on experiments. However, these examples are not intended to restrict 25 the scope of the invention. In the examples, all percentages indicate % by weight.

Example 1:

Commercially-available bleached pulp (trade name "ARAUCO") was made into a 5 % slurry with an acetic acid buffer (pH 6), to 30 which was added SDX enzyme in an amount of 50 U/g (relative to pulp dry matter). The pulp slurry was enzymatically treated at

60°C for 3 hours with stirring, and then de-watered by filtration through a Buchner funnel. The enzymatically-treated pulp thus obtained was then carboxymethyl-etherified to give CMC, according to the CM etherification method mentioned below. The
5 characteristics of the CMC thus obtained herein were compared with those of non-enzymatically treated CMC.

Methods for determining the characteristics of CMC samples are mentioned hereinunder.

CM Etherification of Pulp:

10 A slurry was prepared by stirring pulp with 30 times by weight of 88 % isopropanol. Relative to the amount of glucose units, 1.8 mole of sodium hydroxide was added to form alkali cellulose, followed by 0.8 mole of monochloroacetic acid, and this was reacted at 70 to 80°C for 2.5 hours. After the reaction,
15 the reaction mixture was filtered through a Buchner funnel and washed with an aqueous solution of 75-80 % methanol. This filtration and washing was repeated several times. Then, the residue was dried to obtain a pure CMC.

Filtration Rate:

20 An aqueous solution of 0.5 % CMC sample to be tested was kept at 20°C and applied onto a 200-mesh sieve, whereupon the amount of the filtrate passing through the sieve within 5 minutes was measured using a measuring cylinder.

Viscosity:

25 An aqueous solution of 2 % CMC sample to be tested was kept at 20°C and subjected to viscosimetry using a single cylindrical rotational viscosimeter to determine its viscosity.

Amount of Microgel::

30 An aqueous solution of 0.5 % CMC sample to be tested was kept at 20°C and applied onto a 200-mesh sieve, whereupon the wet

weight of gel remaining on the sieve was measured and represented as % by weight relative to CMC.

	Enzyme-Treated CMC	CMC
Filtration Rate (ml/5 min)	30	18
Viscosity (cps)	1400	1260
Amount of Microgel (%)	4.0	20.5

5 Example 2:

Pulp was enzymatically treated in the same manner as in Example 1, and then modified into CMC according to the alkali cellulose method mentioned below. The characteristics of the CMC thus obtained herein were compared with those of non-
10 enzymatically treated CMC.

Alkali Cellulose Method:

Pulp was soaked in an aqueous solution of 18 % sodium hydroxide. After one or two hours, this was squeezed to remove the excess sodium hydroxide, thereby obtaining an alkali
15 cellulose of 3 times by weight relative to pulp. The resulting alkali cellulose was transferred into a beater. Powder sodium monochloroacetate in an amount of 1.2 to 2.0 mole per mole of anhydrous glucose unit in the pulp was added while beating and stirring. This was further beaten and stirred further for several
20 hours, while keeping the temperature at 10°C or lower, whereby sodium monochloroacetate fully penetrated into the cellulose structure. After this, the resulting mixture was transferred into a reactor, and kept therein at from 70 to 80°C for about 2 hours, with further stirring, to give CMC. The reaction mixture was

filtered and washed several times with an aqueous solution of 75-80 % methanol, and the resulting residue was dried to obtain a pure CMC.

	Enzyme-Treated CMC	CMC
Filtration Rate (ml/5 min)	32	25
Viscosity (cps)	2800	2500
Amount of Microgel (%)	4.5	26.1

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Example 3:

Commercially-available bleached pulp (trade name "ARAUCO") was made into a 15 % slurry with phosphoric acid buffer (pH 8), to which was added SDX enzyme in an amount of 100 U/g (relative
10 to pulp dry matter). With stirring, the pulp slurry was enzymatically treated at 70°C for 5 hours, and then de-watered by filtration through a Buchner funnel. The enzymatically-treated pulp thus obtained was then methyl-etherified to give methyl cellulose, according to the methyl etherification method
15 mentioned below. As a control, pulp that had not been enzymatically treated was methyl-etherified. Its characteristics were compared with those of the enzymatically-treated methyl cellulose.

Methyl Etherification Method:

20 Pulp was soaked in a solution of about 50 % sodium hydroxide, and then squeezed to obtain an alkali cellulose having sodium hydroxide and water in almost the same amount as that of cellulose. To this was added a slight excess of methyl chloride, and this was reacted at 95 to 100°C in an autoclave. After the

reaction, the reaction mixture was washed with hot water on a Buchner funnel and then dried to obtain pure methyl cellulose.

	Enzyme-Treated MC	MC
Filtration Rate (ml/5 min)	28	20
Viscosity (cps)	1100	980

ADVANTAGES OF THE INVENTION

The method of the present invention produces cellulose derivatives with better filterability than conventional cellulose derivatives, while preventing the formation of microgel. The cellulose derivatives thus obtained in the present invention can be effectively used, for example, as thickeners, stabilizers and suspending agents.

Depending on the enzyme used in the method of the invention, the characteristics, such as those mentioned above, of cellulose derivatives obtained may be improved without lowering their viscosity. Thus, the present invention is especially advantageous in this respect.

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u> 4 </u> , lines <u> 33 - 37 </u> to page <u> 5 </u> , lines <u> 1 - 5 </u> .	
B. IDENTIFICATION OF DEPOSIT <div style="float: right; text-align: right;"> Further deposits are identified on an additional sheet <input style="width: 40px; height: 20px; border: 1px solid black;" type="checkbox"/> </div>	
Name of depositary institution National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, Ministry of International Trade and Industry	
Address of depositary institution (<i>including postal code and country</i>) 1-3 Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan	
Date of deposit <div style="text-align: center;">22 December 1993</div>	Accession Number <div style="text-align: center;">FERM BP-4508</div>
C. ADDITIONAL INDICATIONS (<i>leave blank if not applicable</i>) This information is continued on an additional sheet <input style="width: 40px; height: 20px; border: 1px solid black;" type="checkbox"/>	
Until the publication of the mention of grant of a European patent or, where applicable, for twenty years from the date of filing if the application has been refused, withdrawn or deemed withdrawn, a sample of the deposited microorganism is only to be provided to an independent expert nominated by the person requesting the sample (cf. Rule 28(4) EPC). And as far as Australia is concerned, the expert option is likewise requested, reference being had to Regulation 3.25 of Australia Statutory Rules 1991 No 71.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (<i>if the indications are not for all designated States</i>) <div style="height: 40px; border: 1px solid black;"></div>	
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